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conceptual translation product (SEQ ID NO:2) of its longest ORF (GenBank accession number AF 237670). Underlined is a stop codon (TAA) at -6 to -4 that precedes the ATG inframe. Numbers I-XI indicate kinase subdomains as defined by Hanks et al. (25), with invariant and nearly invariant amino acid residues highlighted in black and gray, respectively.--

Page 6, lines 7-13, please replace the paragraph with the following paragraph:

--Fig. 3: Amino acid alignments. (A) Alignment of putative kinase catalytic domain of SOS2 with *Saccharomyces cerevisiae* SNF1, SEQ ID NO:3, (23) and human AMPK kinases, SEQ ID NO:4 (24). Amino acid residues identical in at least two proteins are highlighted in black and conservative substitutions in gray. Mutations that abolish SOS2 autophosphorylation (see Fig. 4) are indicated; first * is K40N,m and second is * G197E, which corresponds to the *sos2-5 allele*. (B) Alignment of the C-terminal portion of SOS2 with the regulatory domains of *Schizosaccharomyces pombe* (yCHK1, SEQ ID NO:5) and human CHK1 (hCHK1, SEQ ID NO:6) kinases (27).--

Page 13, prenumbered lines 4-24, please replace the paragraph with the following paragraph:

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--Protein Expression. To produce bacterially expressed recombinant proteins, the coding region of SOS2, SOS2(K40N), and SOS2(G197E) cDNAs were amplified by PCR with primers harboring restriction sites, cloned in frame into BamHI-EcoRI of pGEX-2TK (Amersham Pharmacia), and transformed into Escherichia coli BL21 DE3 cells (Amersham Pharmacia). Mutations K40N and G197E in the SOS2 protein were created by site-directed mutagenesis. For glutathione S-transferase (GST)-SOS2(K40N), primer pairs 5'-GCGGATCCATGACAAAGAAAATGAGAAGAGTGGGC (SEQ ID NO:7) and 5'-

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ATTGTACTCTTAGCCATAATGTTGATGGCT (SEQ ID NO:8) were used for the first PCR, and 5'-GCGAATTCTTAAGTTGGGATCAAAACGTGATTGTTCTG (SEQ ID NO:9) and 5'-GTGATAATGTAGCCATCAACATTATGGCTA (SEQ ID NO:10) were used for the second PCR. For the mutant protein GST-SOS2 (G197E), primer pairs 5'-GCGGATCCATGACAAAGAAAATGAGAAGAGTGGGC (SEQ ID NO:11) and 5'-ATATAACGAAAAGAATAACCTCGCAAGACC (SEQ ID NO:12) were used for the first reaction and 5'-GCTGATATTTGGTCTTGCGAGGTTATTCTT (SEQ ID NO:13) and 5'-GCGAATTCTTAAGTTGGGATCAAAACGTGATTGTTCTG (SEQ ID NO:14) were used for the second reaction. The final amplification was done with 5'-GCGGATCCATGACAAAGAAAATGAGAAGAGTGGGC (SEQ ID NO:15) and 5'-GCGAATTCTTAAGTTGGGATCAAAACGTGATTGTTCTG (SEQ ID NO:16) on both templates. The final constructs were confirmed by sequencing *E. coli* cultures were induced with 0.5 mM isopropyl β-D-thiogalactoside, and recombinant proteins were affinity-purified from bacterial lysates with glutathione-Sepharose beads (Amersham Pharmacia).--

Page 25 (Abstract), after the last line, beginning on a new page, please insert the attached Sequence Listing.

REMARKS

Claims 1-42 are pending in the present application.

Applicants have now submitted a Sequence Listing and a corresponding computer-readable Sequence Listing. The sequence information recorded in the corresponding computer-readable Sequence Listing is identical to the paper copy of the Sequence Listing. Support for all of the sequences listed in the Sequence Listing is found in the present